

ASST Ovest Milanese (West Milan Local Healthcare Authority) Department of Services and Diagnostic Imaging	INFORMATION ON HOW TO COLLECT STOOL SAMPLES	MAC221-B Rev4 Page 1 of 2
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		VERIFICATION AND APPROVAL	
Revision	Date	Office	Approved by
0	2021	Director of the Microbiology Unit and Head of the Analysis Laboratory Unit Director of the Pathological Anatomy Unit	Dr Pierangelo Clerici Dr Paolo Fociani
1	June 2022	Director of the Microbiology Unit Acting Director of the Analysis Laboratory Unit Director of the Pathological Anatomy Unit	Dr Pierangelo Clerici Dr Sergio Finazzi Dr Paolo Fociani
2	November 2022	Director of the Microbiology Unit Acting Director of the Analysis Laboratory Unit Director of the Pathological Anatomy Unit	Dr Pierangelo Clerici Dr Sergio Finazzi Dr Paolo Fociani
3	November 2024	Acting Director of the Microbiology Department Director of the Analysis Laboratory Department Director of the Pathological Anatomy Department	Dr Bianca Osnaghi Dr Sergio Finazzi Dr Paolo Fociani
4	February 2025	Acting Director of the Microbiology Department Director of the Analysis Laboratory Department	Dr Bianca Osnaghi Dr Sergio Finazzi

TYPES OF ANALYSIS ON FAECES

GENERAL INSTRUCTIONS ON HOW TO COLLECT STOOL

1. **Collect** stool on a dry surface
2. **DO NOT** urinate on the sample during collection
3. **DO NOT** pass stool into the toilet bowl.

☐ **FAECES DOSAGE ELASTASIS, CALPROTECTIN**

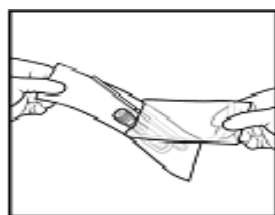
1. Collect a small amount of stool with the scoop attached to the lid, place it in the bottle and close carefully.
2. Deliver on the first available working day, storing meanwhile the sample in the refrigerator
3. If the investigation is requested **on multiple samples**: collect from different discharges, store the samples in the refrigerator for a maximum of three days and deliver on the first available working day.

☐ **OCCULT BLOOD DETECTION**

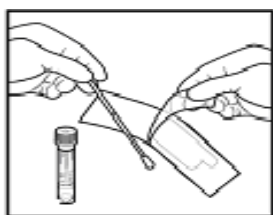
1. For female users, do not perform the test during menstruation
2. Unscrew and remove the cap from the tube
2. Swipe the stick over the stool several times, horizontally and vertically.
3. Reinsert the rod into the tube, screw it back on and shake
4. Deliver on the first available working day, storing meanwhile the sample in the refrigerator
5. If the investigation is requested **on multiple samples**: collect from different discharges, store the samples in the refrigerator for a maximum of three days and deliver on the first available working day

☐ **STOOL CULTURE (Salmonella, Shigella, Campylobacter)**

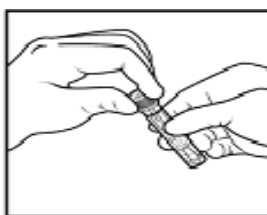
1. Collect a small portion of stool with the swab following the procedure shown below



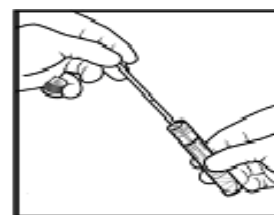
Open the package



Collect a small portion of stool with the swab



Open the test tube



Insert the swab.
Break the rod
by the red edge and close
the test tube

2. After collection, store the container in the refrigerator at 2-8° C.
3. If the investigation is requested **on multiple samples**: collect from different discharges, store the samples in the refrigerator for a maximum of 48 hours and deliver on the first available working day.

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❑ SEARCH FOR TOXINS, ANTIGENS AND INTESTINAL VIRUSES (*Clostridium difficile*, Rotavirus, Adenovirus, *Giardia lamblia*, *Entamoeba histolytica*, *Cryptosporidium parvum*, *Helicobacter Pylori*)

1. **DO NOT take**, for a week before and during collection, interfering substances such as laxatives, barium, bismuth, mineral oils, antibiotics, or antidiarrhoeals.
2. For the Helicobacter Pylori Antigen search, antacids and proton pump inhibitors must not be taken for at least two weeks before the sample collection. If you are currently undergoing therapy, DO NOT interrupt it and contact your GP before performing the test.
3. Collect a small amount of stool with the scoop attached to the lid, place it in the bottle and close carefully.
4. Deliver within the first hours of collection.
5. If the investigation is requested **on multiple samples**: collect the samples on consecutive days, delivering each collection daily as explained under point 4.

❑ PARASITE DETECTION

1. **DO NOT take** interfering substances such as laxatives, barium, bismuth, mineral oils, antibiotics or antidiarrhoeals.
2. **Bottle with preservative (green cap)**: collect a significant portion of faeces (preferably from bloody or mucoid areas) with the scoop attached to the lid and introduce a quantity of material sufficient to reach the red line on the bottle, without exceeding it.
Close and shake vigorously until the material is well homogenized. Store this container at room temperature both before and after collection (do not put it in the refrigerator!)
NB: the preservative liquid in the green container is poisonous and irritating. It must not be ingested or brought into contact with the skin and mucous membranes. In case of contact with the skin, wash thoroughly with soap and water. In case of ingestion, consult your doctor.
3. **Container without liquid (red cap)**: collect a significant sample of faeces with the scoop attached to the lid, and place it in the container **without exceeding half**. Store the container at room temperature.
4. If **adult worms or proglottids** are found, they must be placed in the container for the collection of fresh faeces (red cap) and never in the green containers (with preservative liquid)
5. If the investigation is requested **on 2 samples**: collect the samples on consecutive days using a green container on the first day and another green container and the red cap on the second day.
6. If the investigation is requested **on 3 samples**: collect the samples on consecutive days using the two green containers on the first two days and the container with the red cap on the third day.
7. Deliver all samples at the same time, together with the duly completed medical history form (MUC08) duly filled in.

❑ SCOTCH-TEST (Pinworm search, *Enterobius vermicularis*)

1. A slide and a Petri dish are used for collection, together with transparent scotch tape (adhesive tape not supplied by the laboratory).
2. The sample must be taken in the morning, upon waking up.
3. Before sampling, do not wash the anal region and do not use any kind of ointment.
4. Make the adhesive part of the scotch tape adhere to the anal region without the scotch tape forming creases, leaving it for about 15 seconds.
5. Remove the scotch tape and place it on the slide well stretched **avoiding the formation of creases and bubbles**.
6. Place the slide with the scotch tape adhering to the Petri dish.
7. If the investigation is requested **on multiple samples**: collect the samples on consecutive days, storing at room temperature and deliver them together on the first available working day.

❑ STOOL FOR KOCH'S BACILLUS DETECTION (*Mycobacteria*)

1. Collect the stool samples, using a jar with a shovel.
2. Deliver the samples within the first hours of collection.
3. If the investigation is requested **on multiple samples**, collect on consecutive days, delivering each sample as explained under point 2.